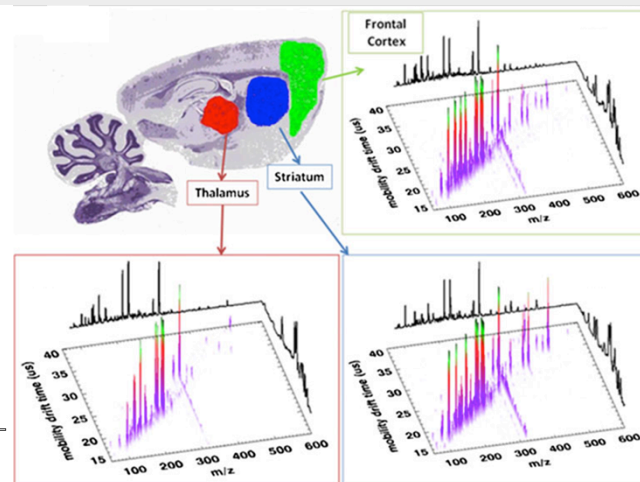
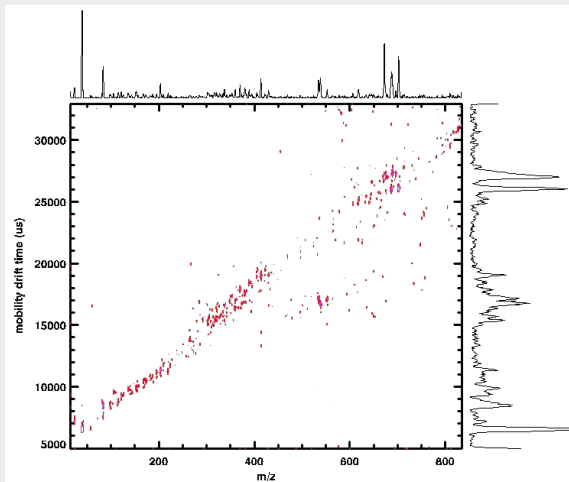
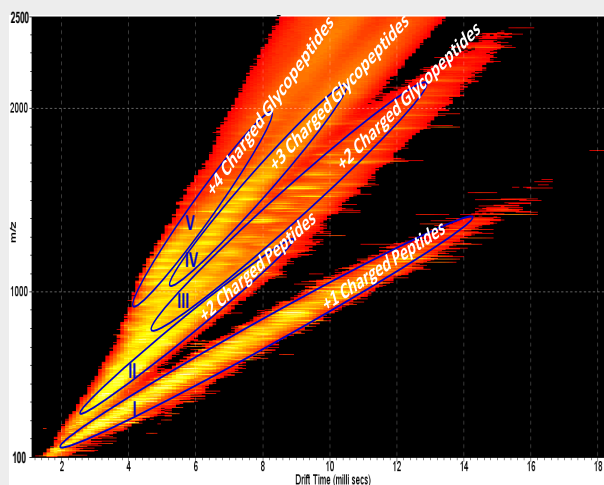


Ion Mobility Workshop

Future of Ion Mobility Mass Spectrometry

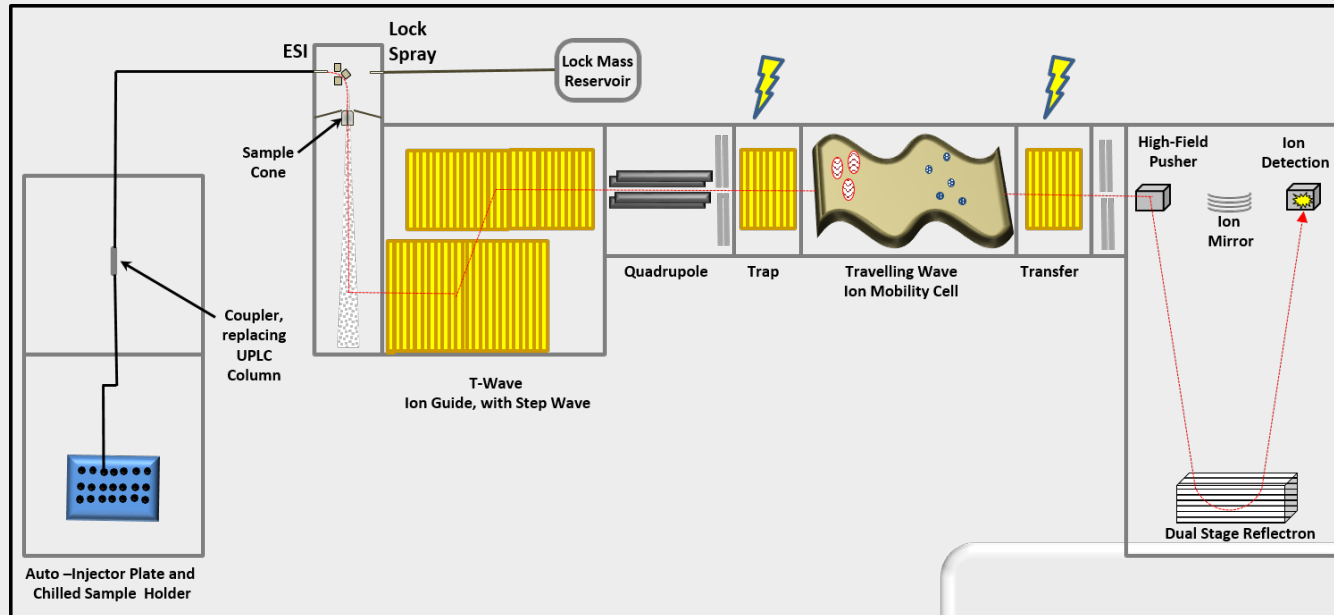
--- To Biological Problems

Herb Hill
Department of Chemistry
Washington State University

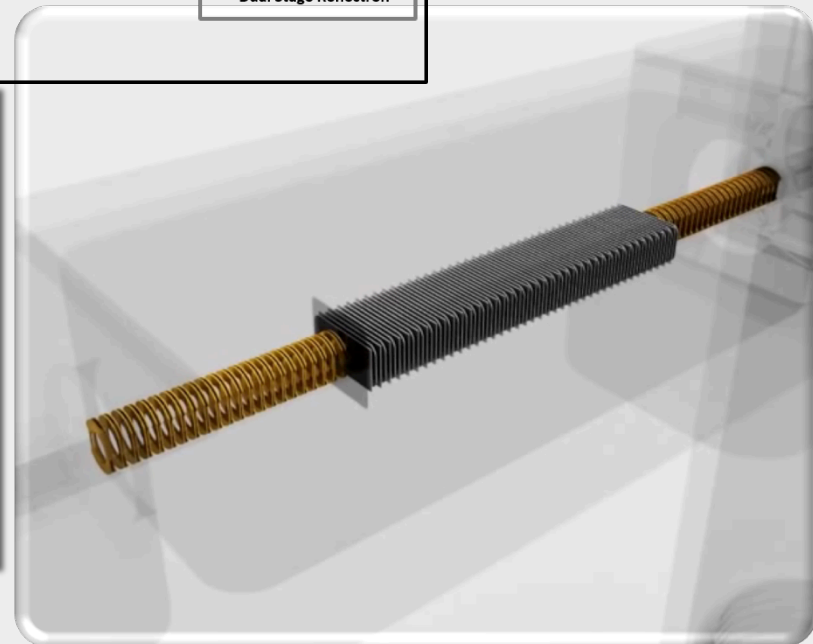




Synapt G2-S HDMS



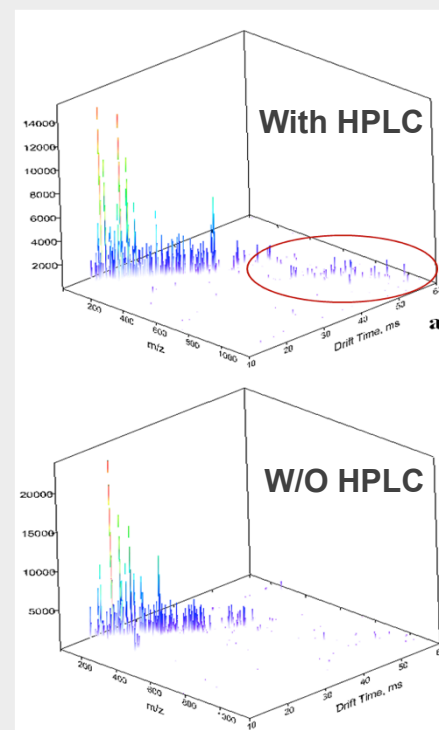
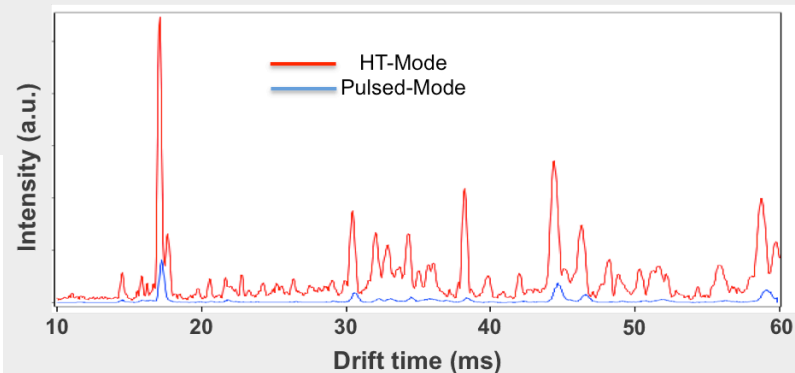
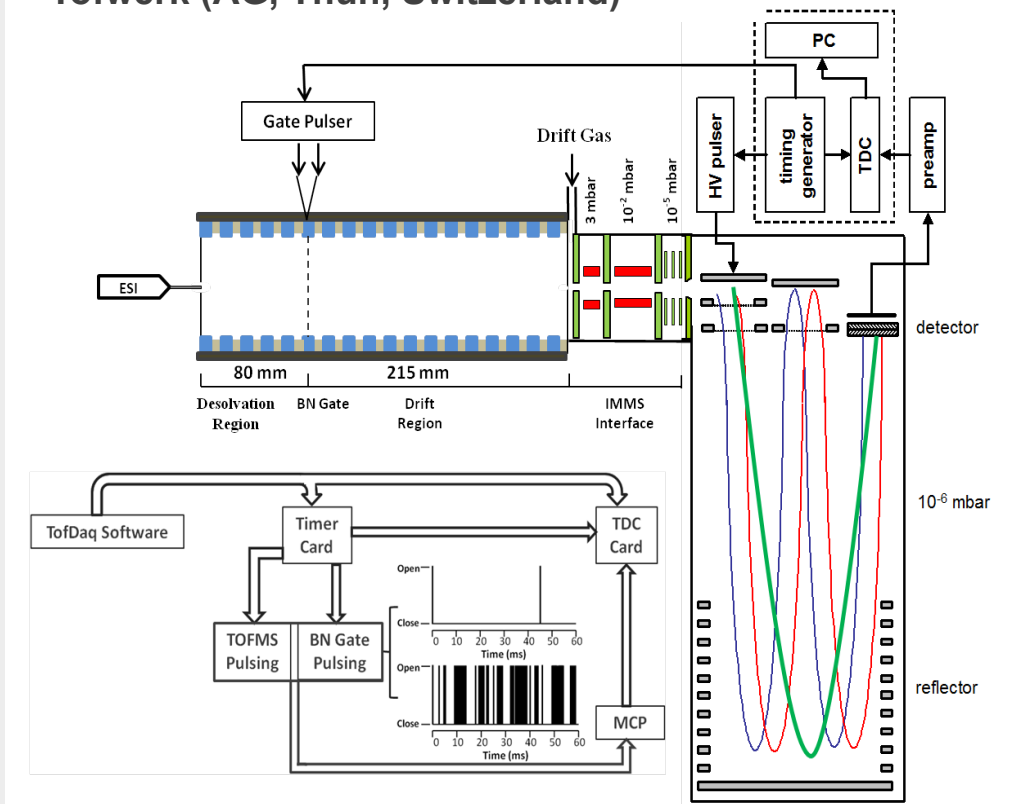
- **Synapt G2-S HDMS system**
 - Operated in the Mobility-Mass mode with TOF section in “V”, or “resolution” configuration
 - TWIMMS separates ions, as they “surf” along a pulsed, time varying electric field, according to size and charge.
 - Direct infusion was achieved by use of a Waters ACQUITY UPLC
 - In place of column a VALCO coupler is placed in line to allow direct infusion.
 - Reduced pressure, 3-4 mbar Nitrogen
 - Moving electric field
 - Resolving power is relatively low (30-40)
 - Fewer Collisions with Drift Gas



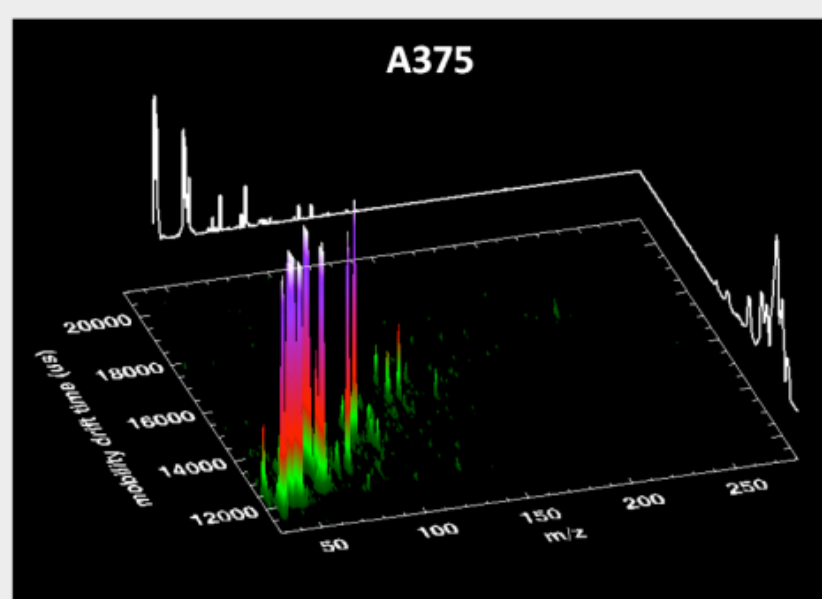
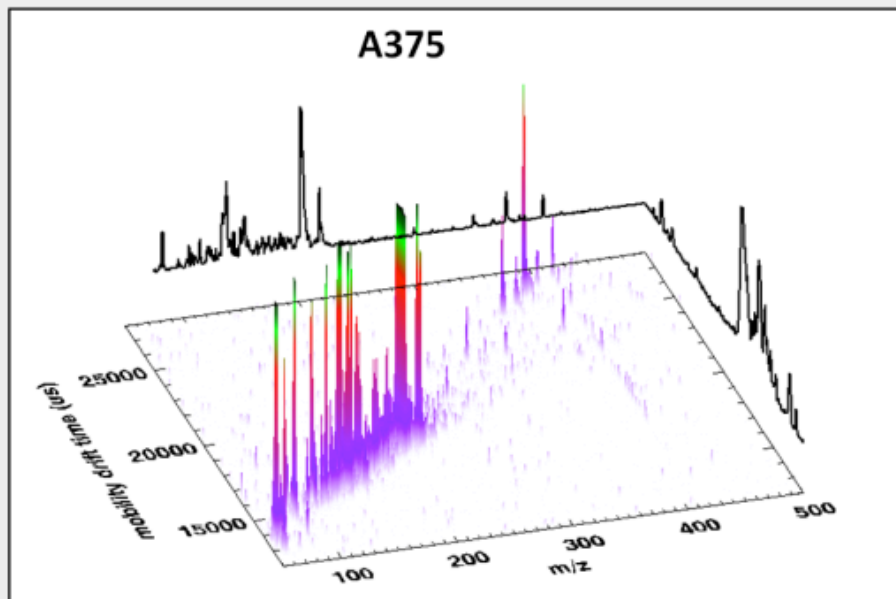
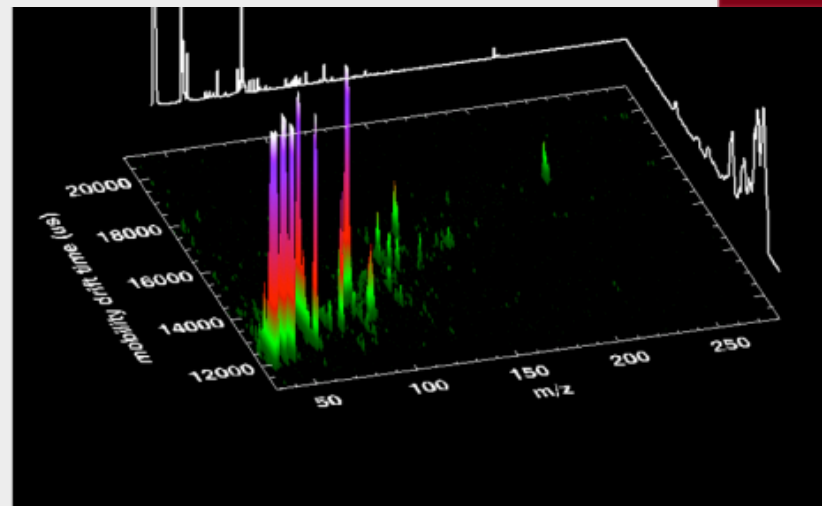
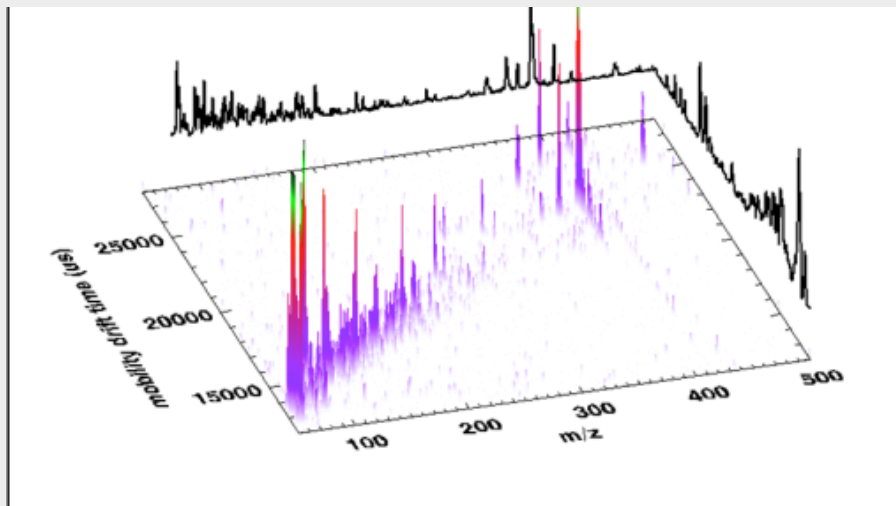
High Throughput Metabolomics analysis---HT-apIMMS



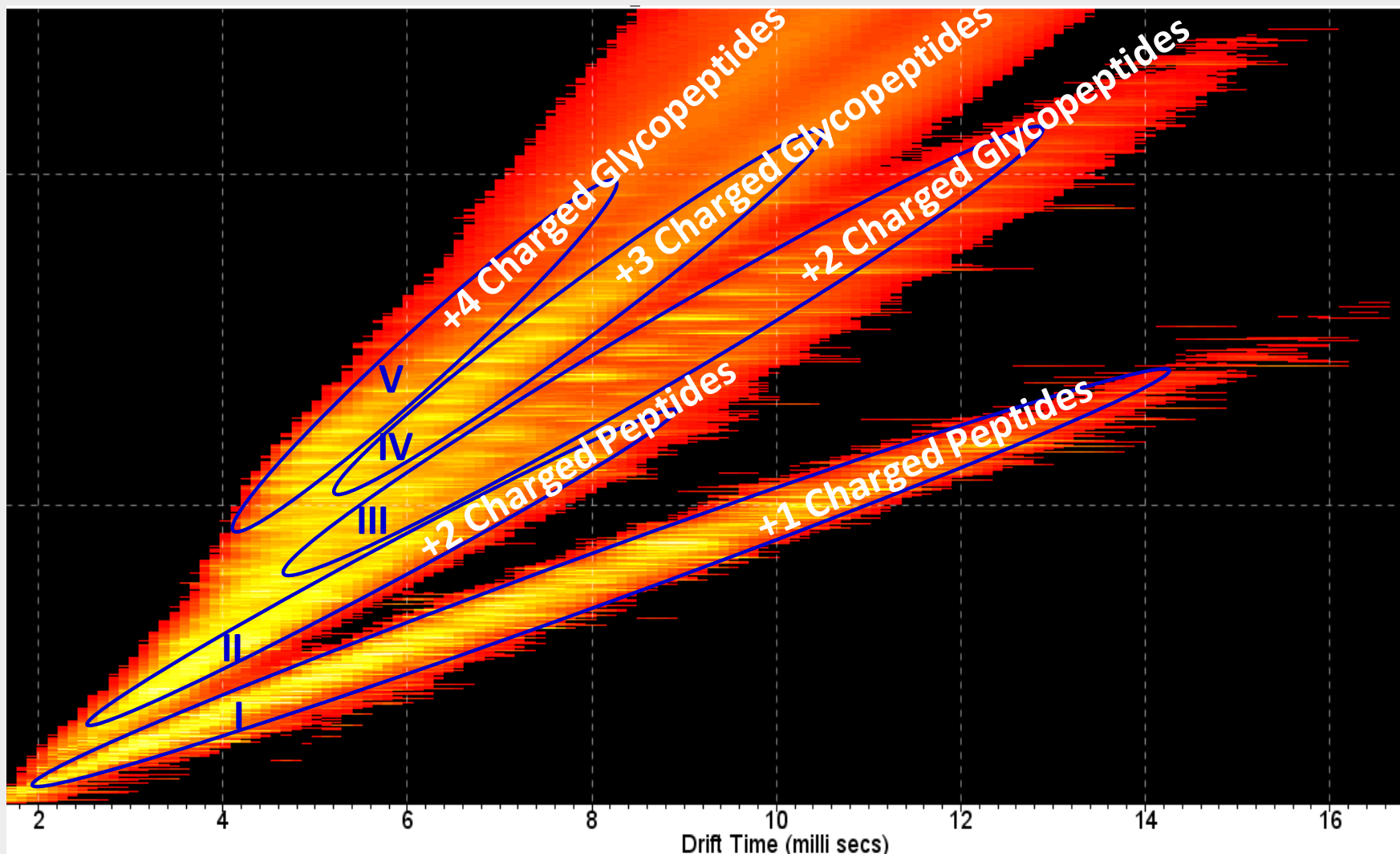
Tofwerk (AG, Thun, Switzerland)



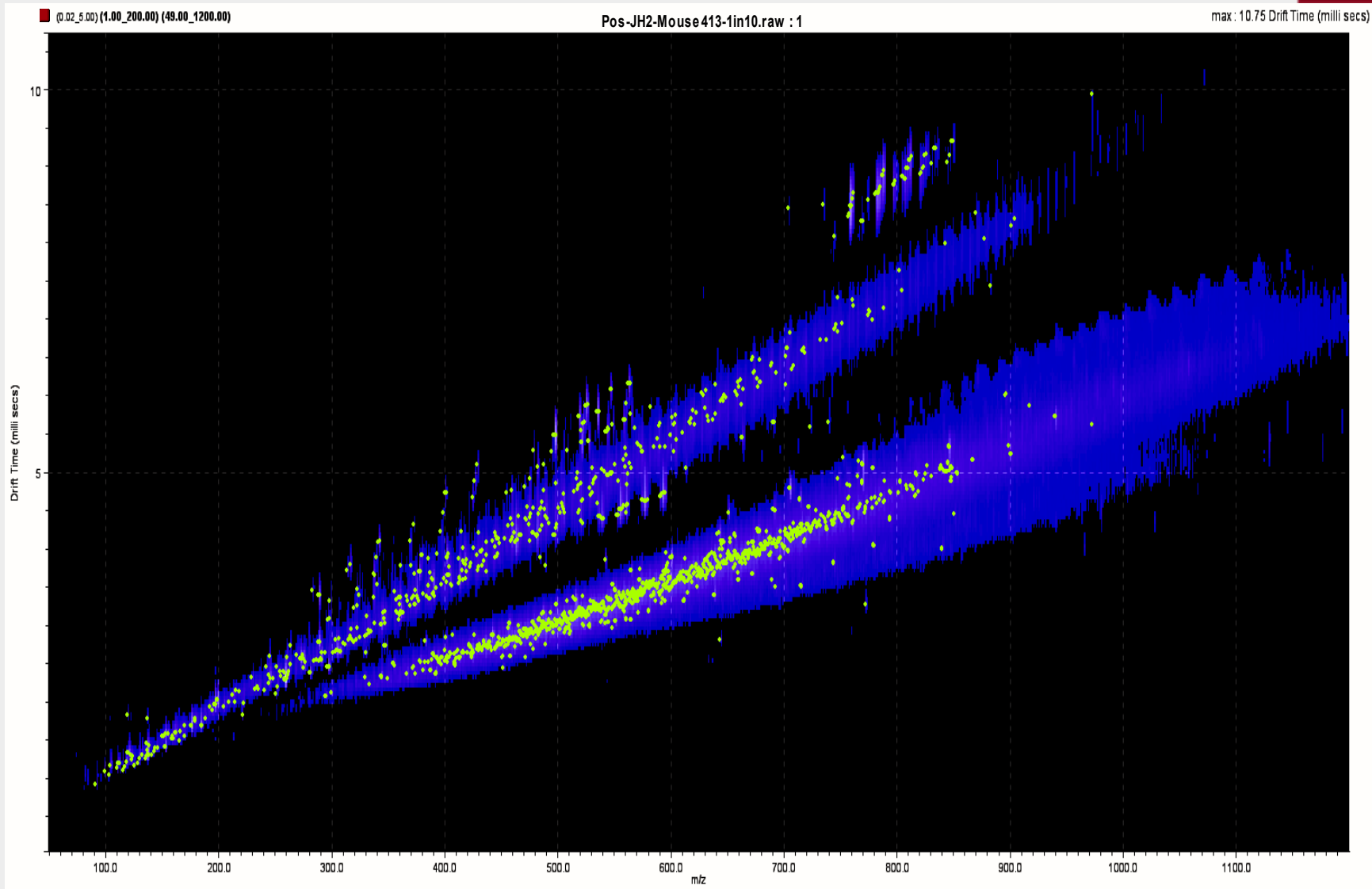
Melanoma Cancer Cell



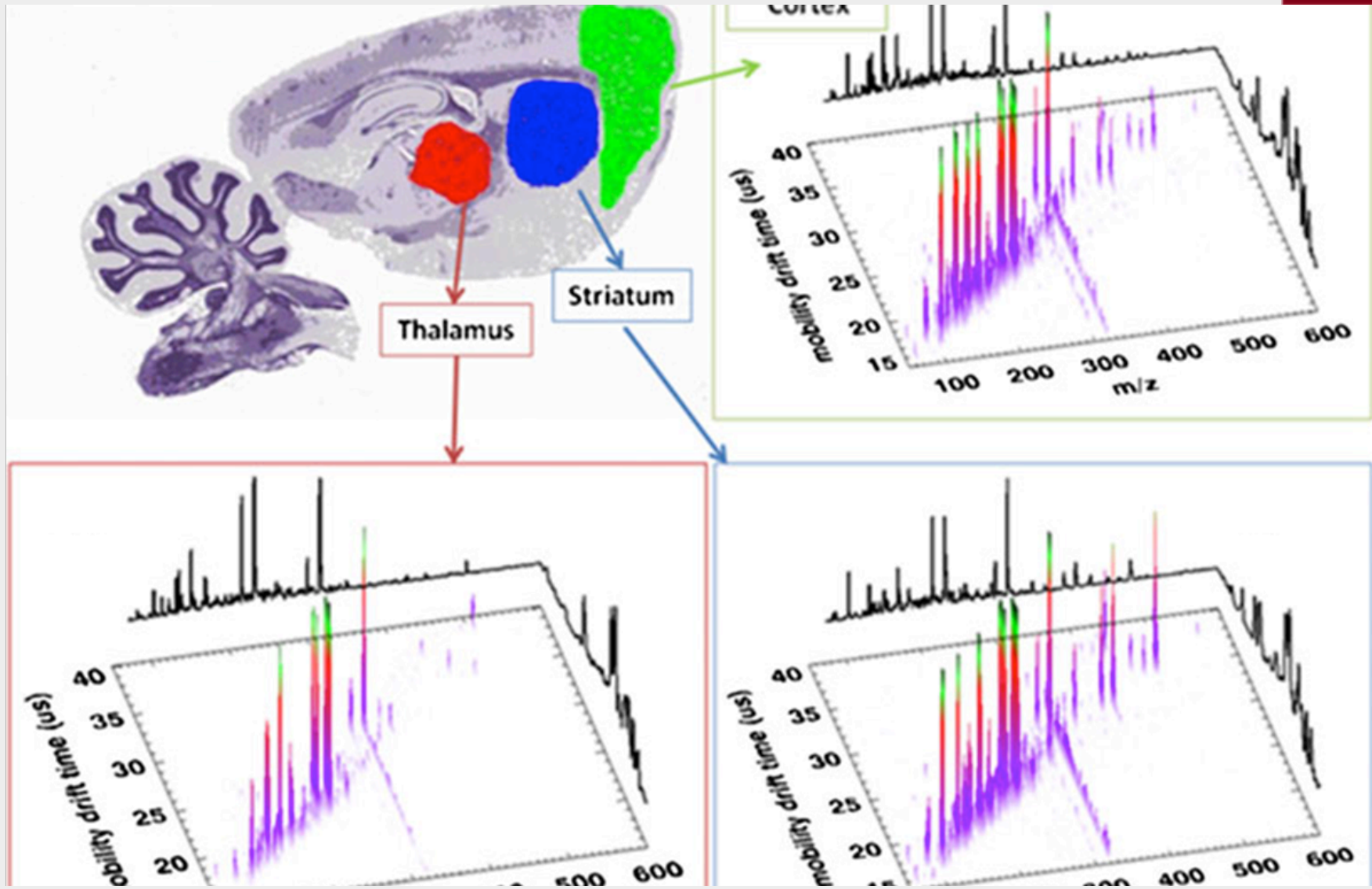
Glycopeptides



Colorectal Cancer Tissue



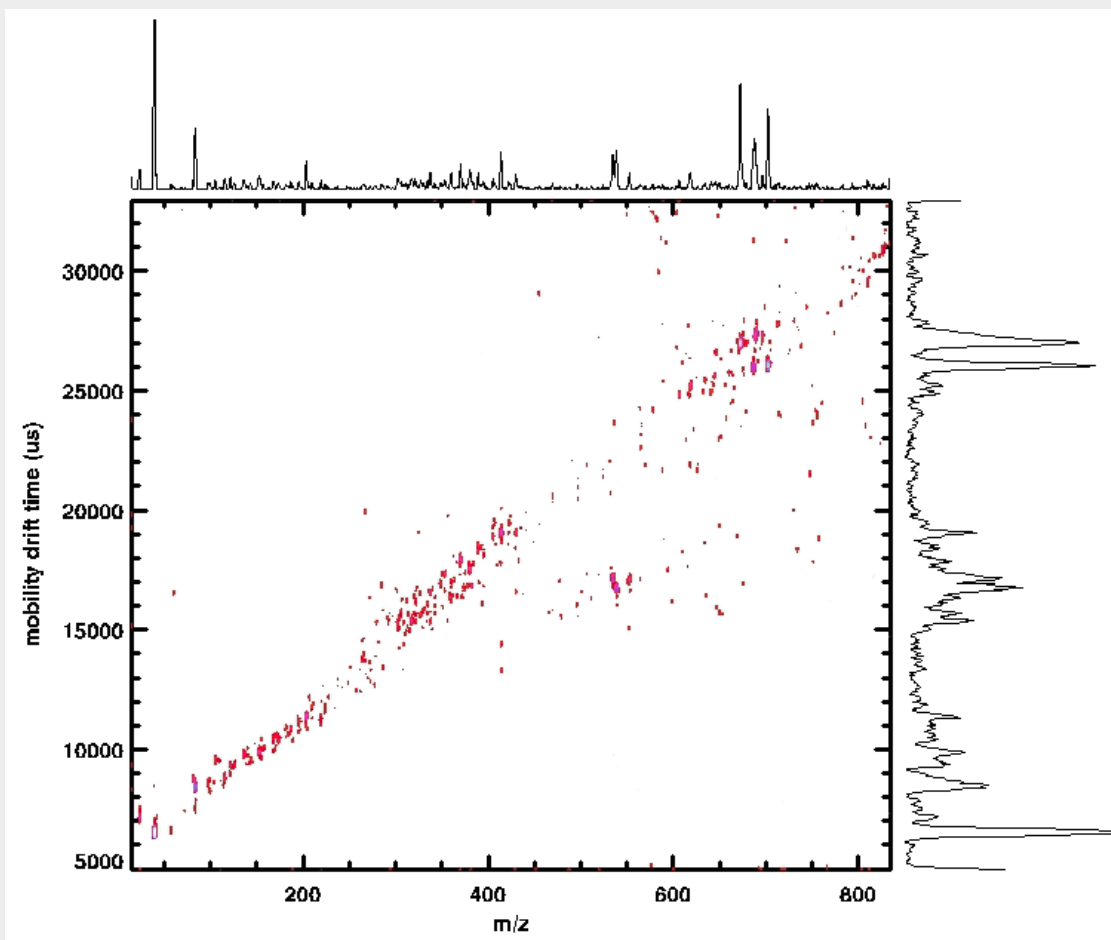
Brain Tissue



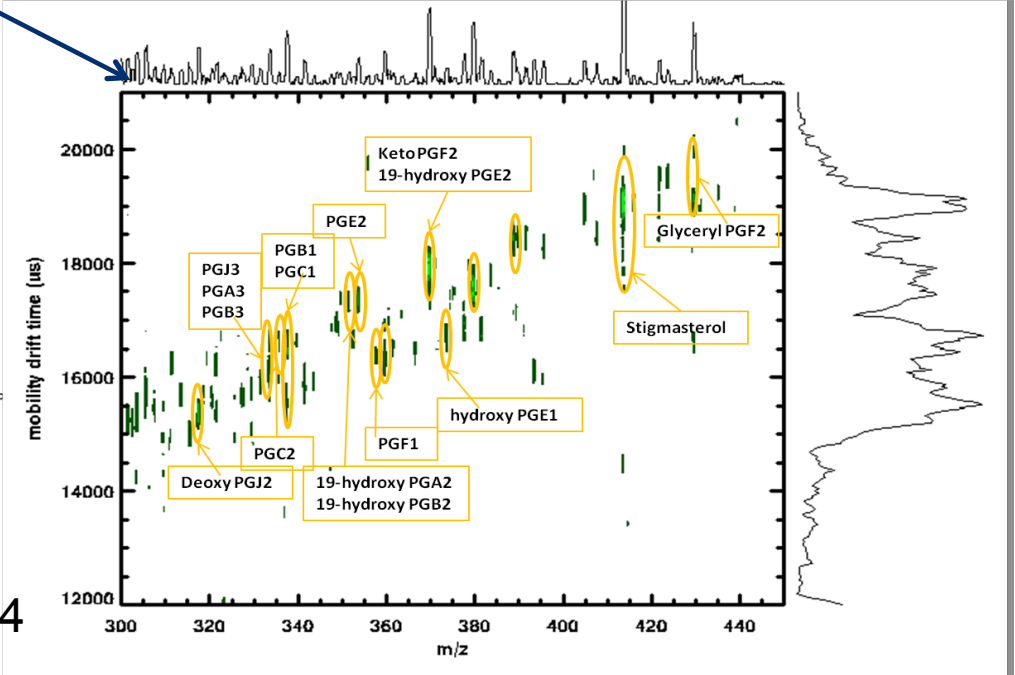
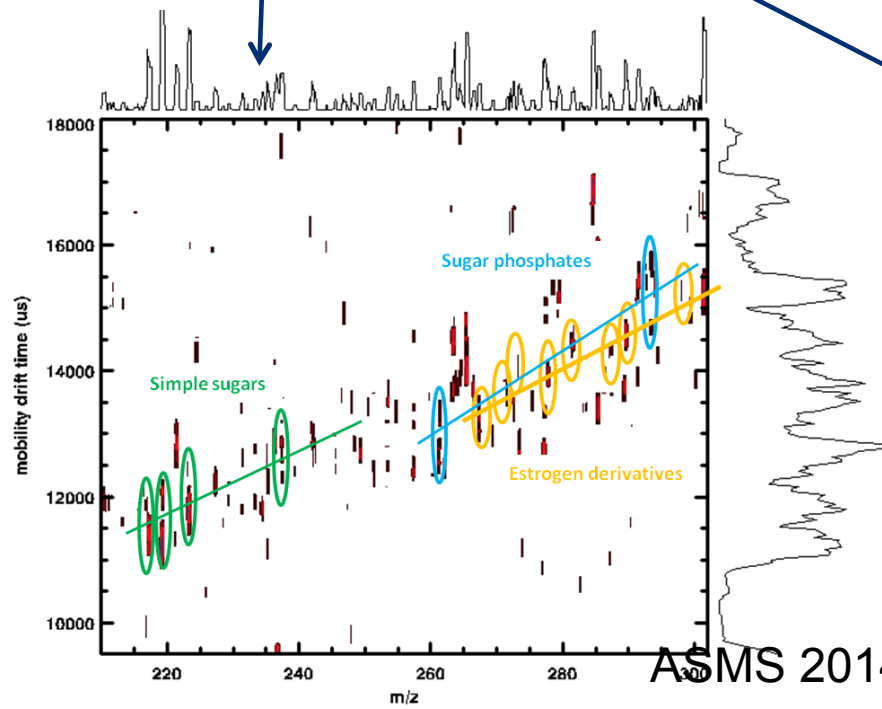
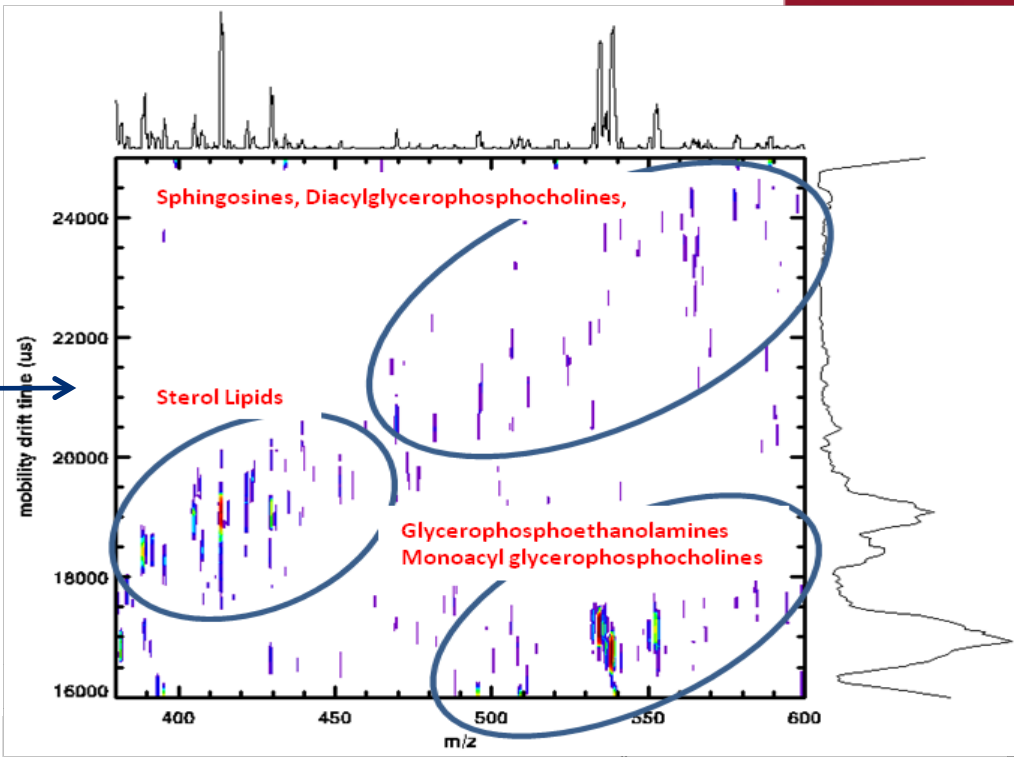
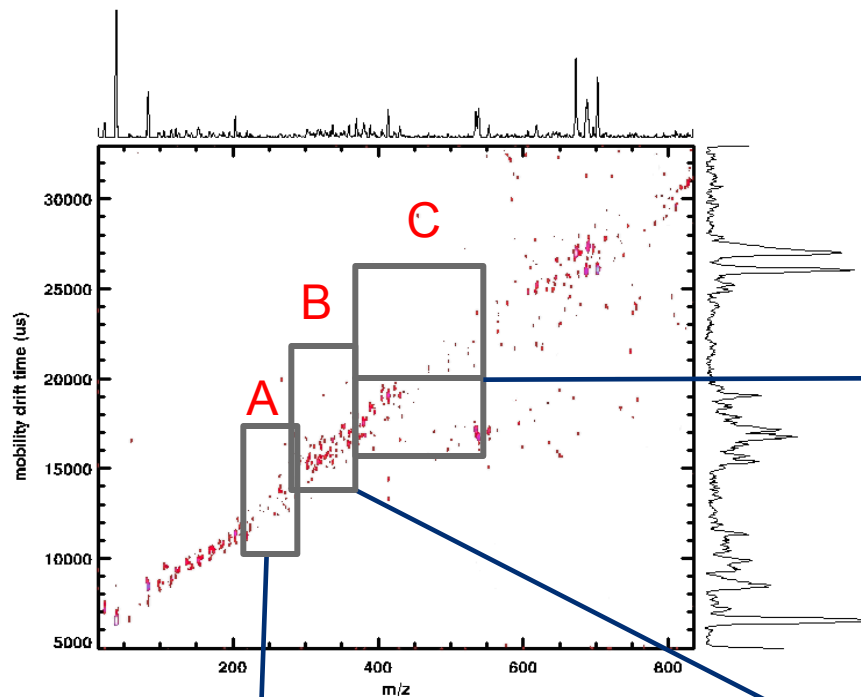
Metabolome of Human Blood



Few Drops of Blood
Extracted with MeOH/Water
Centrifuged
ESI-IMS-TOF
30 minutes
10 nM to 10 μ M
200 isobars



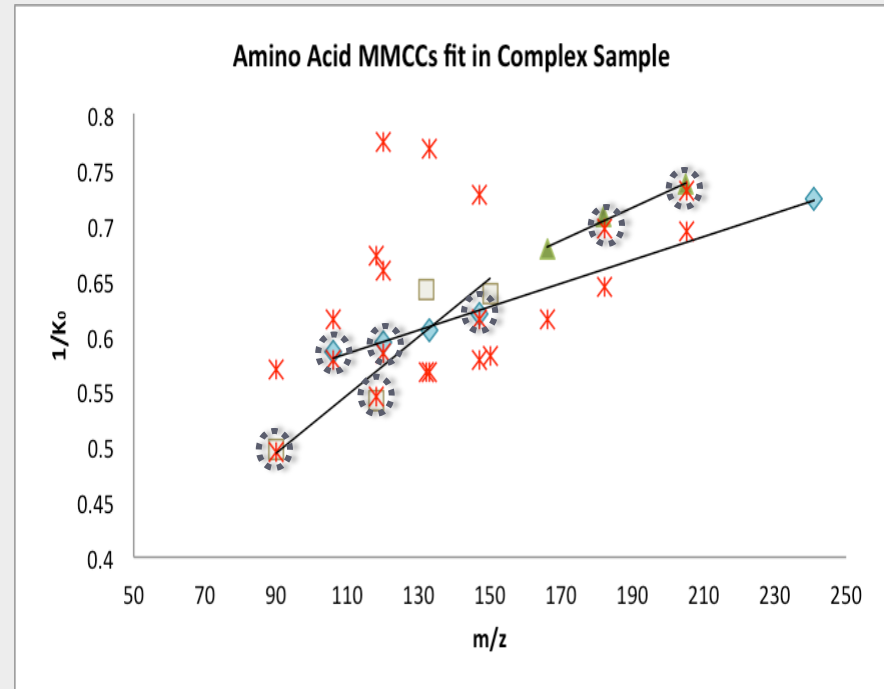
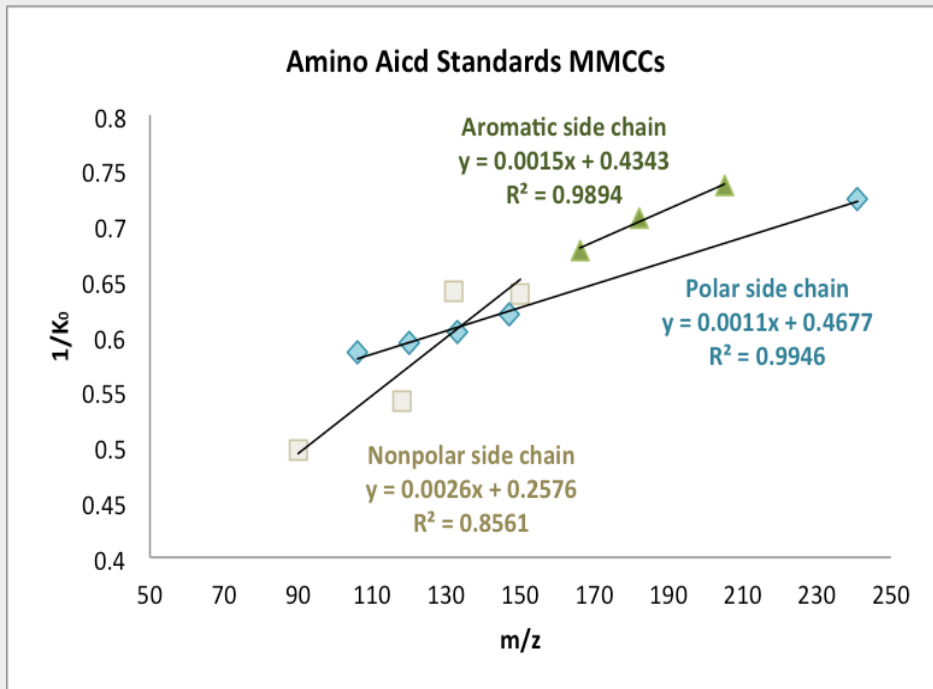
Metabolic profiling of human blood by high-resolution ion mobility mass spectrometry (IM-MS), P. Dwivedi, J. A. Schultz, H. H. Hill Jr, *Int. J. Mass Spectrom.* 298 (2010) 78-90.



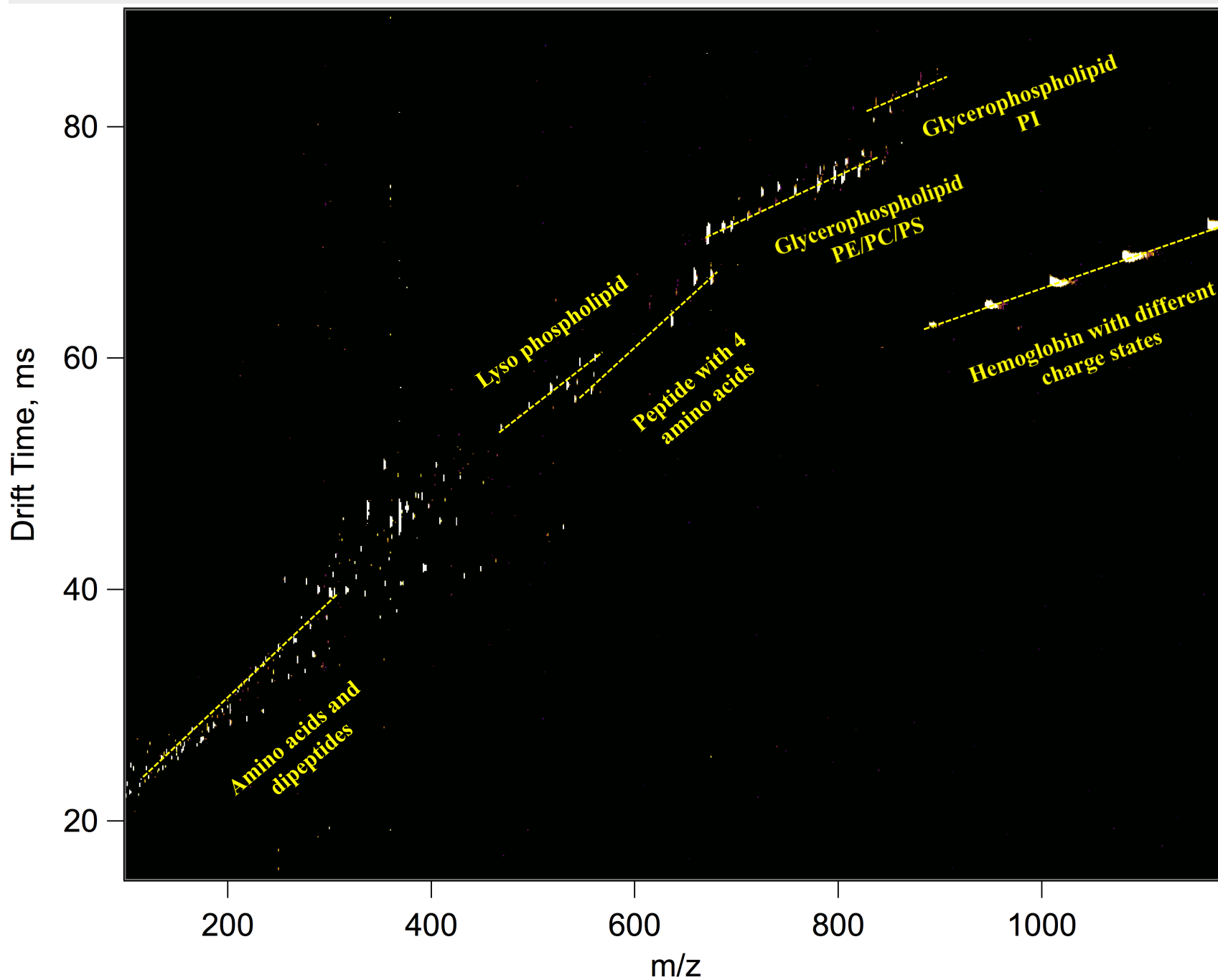
ASMS 2014

Structural Elucidation of Unknowns

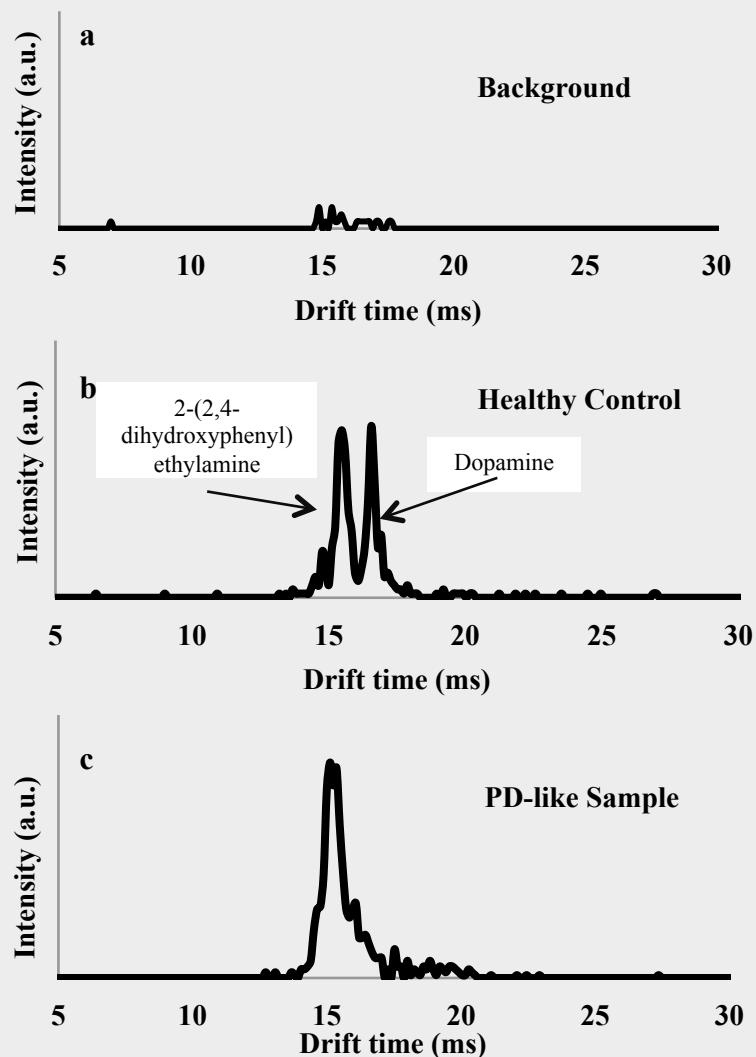
---Mobility Mass Correlation Curves (MMCCs)



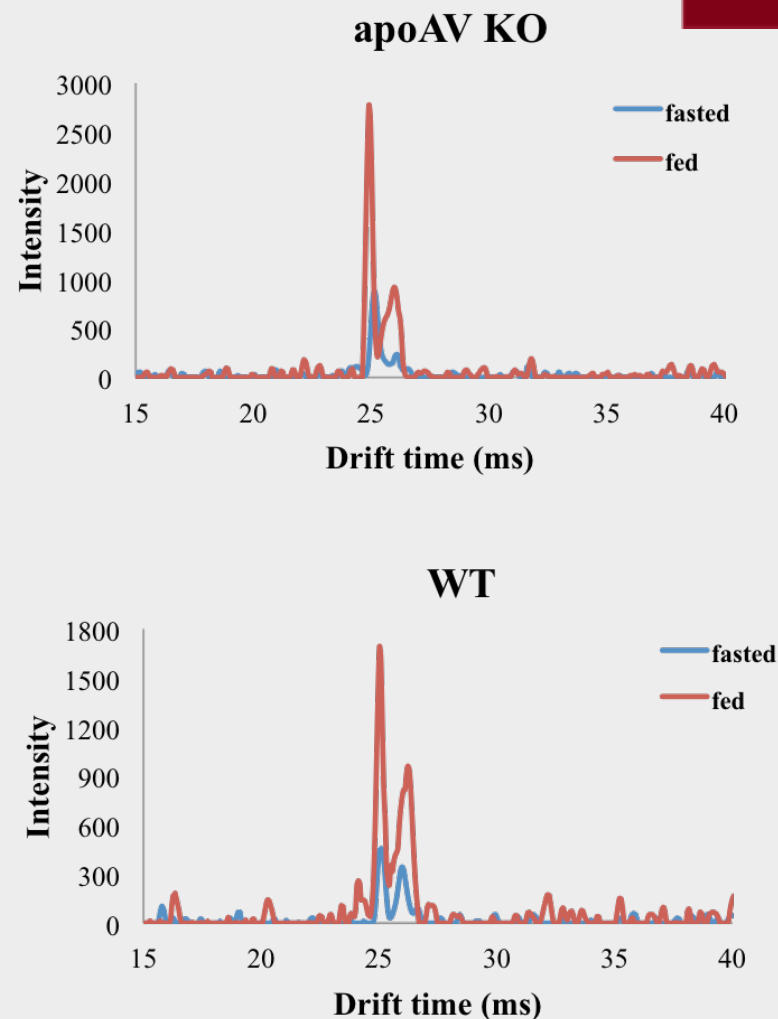
Identification of amino acids in brain tissue (striatum) using MMCCs obtained by standard analysis.



Isomeric Separations with High Resolving Power



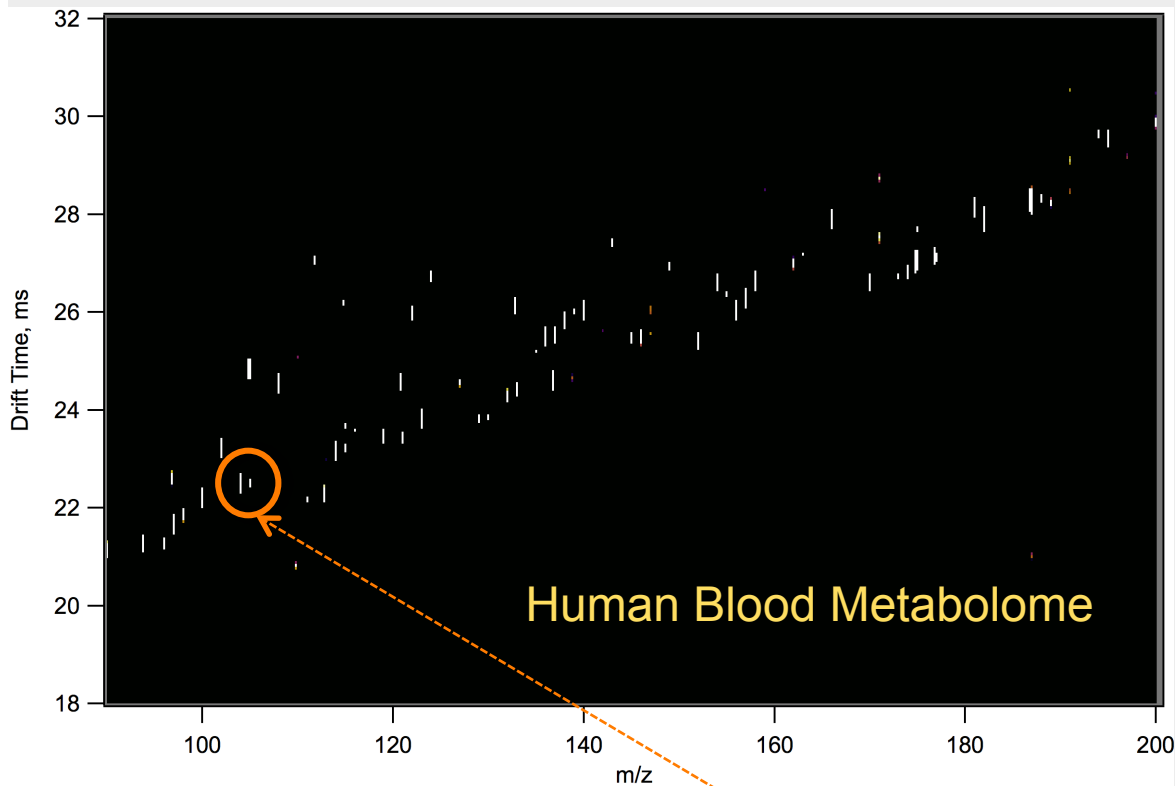
Detection of dopamine and its isomer in striatum from healthy control and PD-like sample



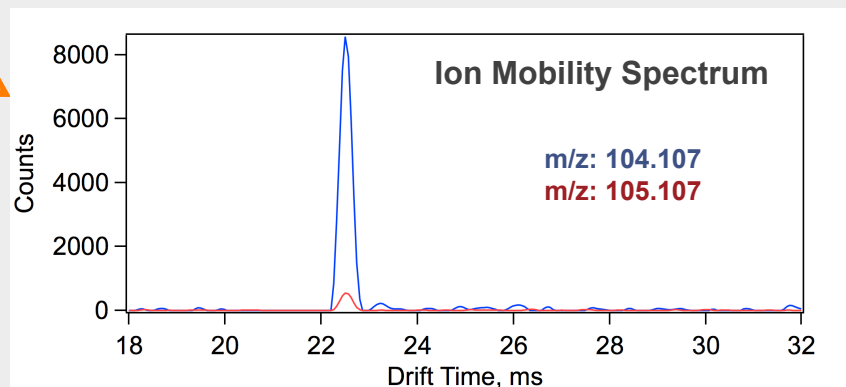
Detection of glucose and fructose in plasma fluids from apoAV KO mice and WT mice

Structural Elucidation of Unknowns

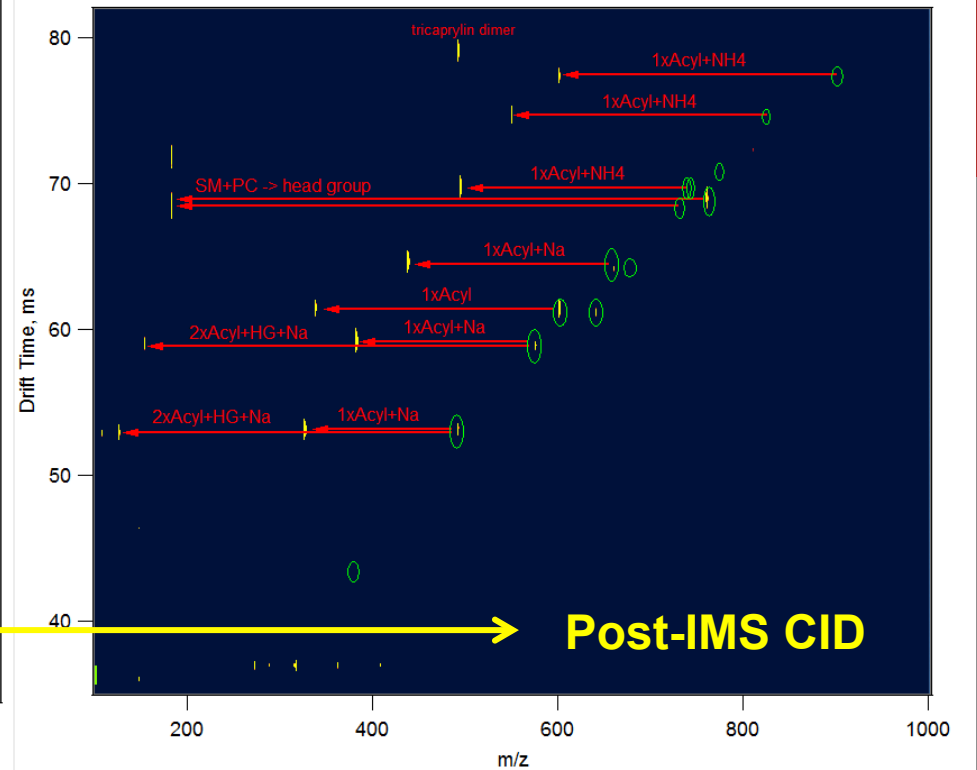
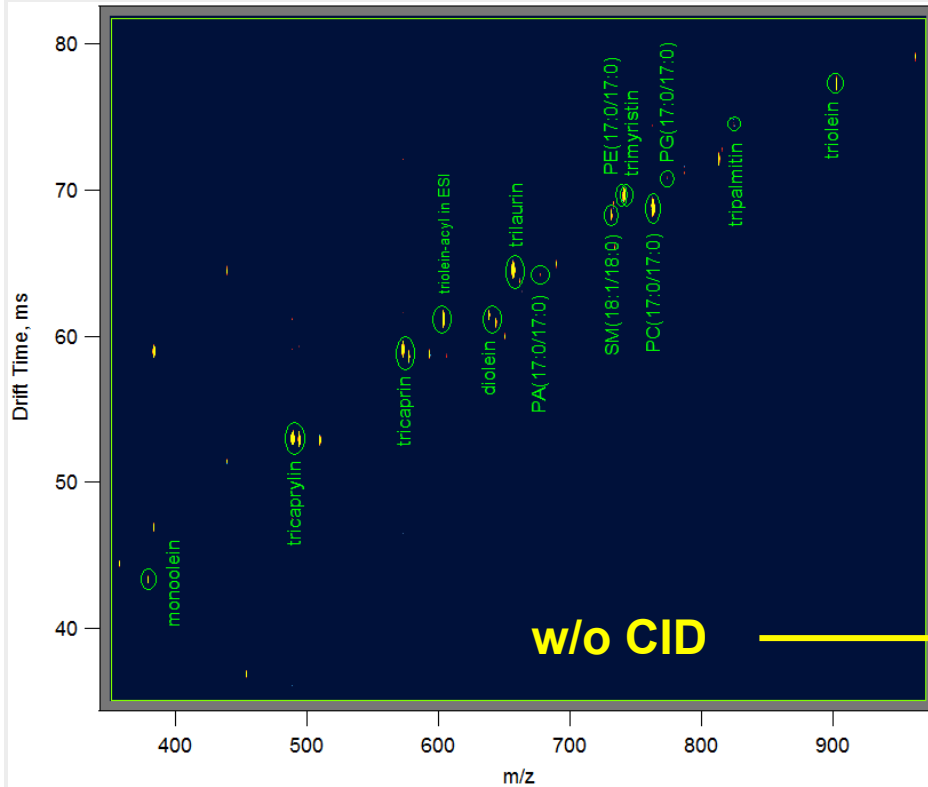
---Isotopic Ratio Analysis



Choline
Isotopic ratio 100:5.5



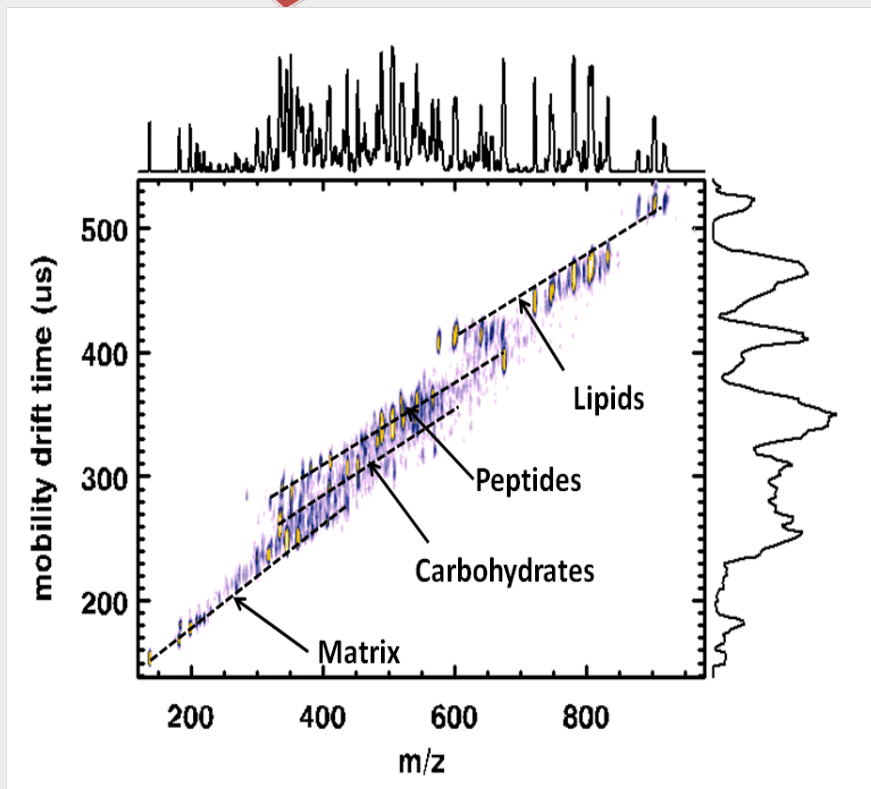
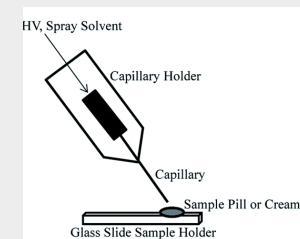
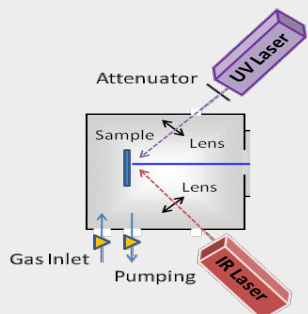
Structural Elucidation of Unknowns ---Fragmentation Analysis



IMMS coupled with Surface Ionization

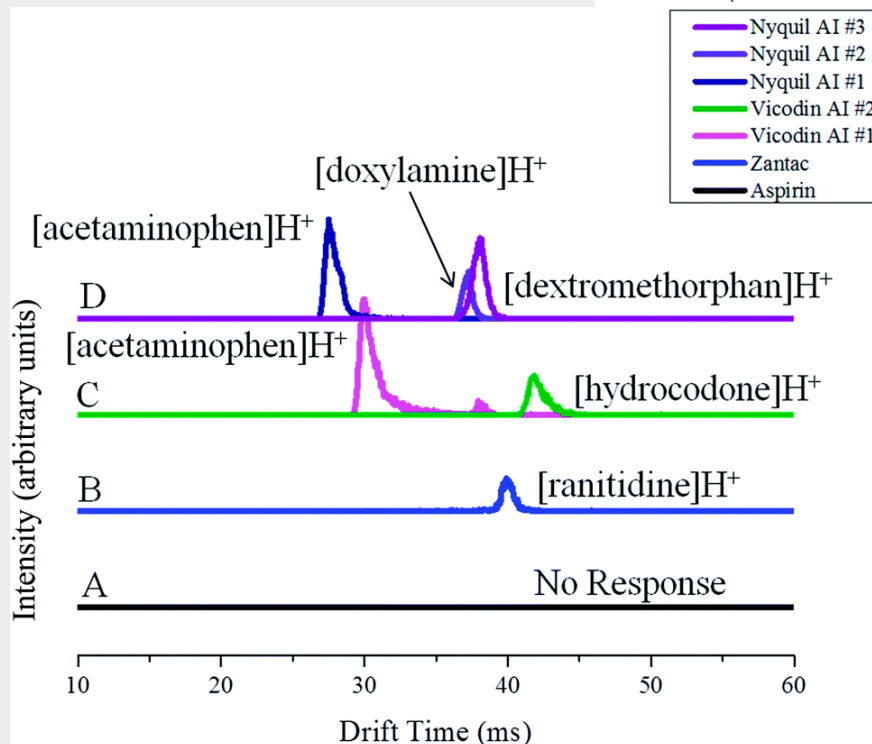
---Minimize Sample Preparation

---Imaging



MALDI-IMMS of mice lymph fluid

Kaplan et al. *Int. J. Ion Mobil. Spec.* 2013, 16, 177-184.



DESI-IMMS of drugs

Roscioli et al. *Analyst.* 2014, 139, 1740-1750.

IMMS Database with Accurate K_0 Measurement



Standards measured at 30°C, 477 V/cm, water < 1ppm_v

Standard	K_0	Literature K_0	Reference
PDO (- mode)	1.784 ± 0.001	--	--
PDO (+ mode) Monomer	1.853 ± 0.002	2.03 (200°C); 2.04 (200°C)	1, 2
PDO (+ mode) Dimer	1.476 ± 0.001	--	--
2,4-Lutidine Monomer	1.867 ± 0.002	1.84-2.07 (78°C-250°C)	3
2,4-Lutidine dimer	1.404 ± 0.001	1.37-1.38 (37°C-38°C)	3
DtBP	1.480 ± 0.002	1.43-1.44 (37°C-250°C); 1.42 (25°C)	3, 4
DMMP Monomer	1.805 ± 0.002	1.80-2.05 (37°C-250°C); 1.74-1.80 (25°C)	3, 4
DMMP Dimer	1.429 ± 0.001	1.39-1.40 (95°C-150°C); 1.38-1.39 (25°C)	3, 4

1. Karpas et al. *Int. J. Mass Spectrom. Ion Processes*. 1991, 107, 435-440.
2. Berant et al. *J. Am. Chem. Soc.* 1989, 111, 3819-3824.

3. Eiceman et al. *Anal. Chim. Acta.* 2003, 493, 185-194.
4. Viitanen et al. *Talanta.* 2008, 76, 1218-1223.

IMMS Database with Collision Cross Sections (CCS)



$$\frac{\Omega}{z} = \frac{3}{4} \left(\frac{e}{\mu v_T K_0 N_0} \right)$$

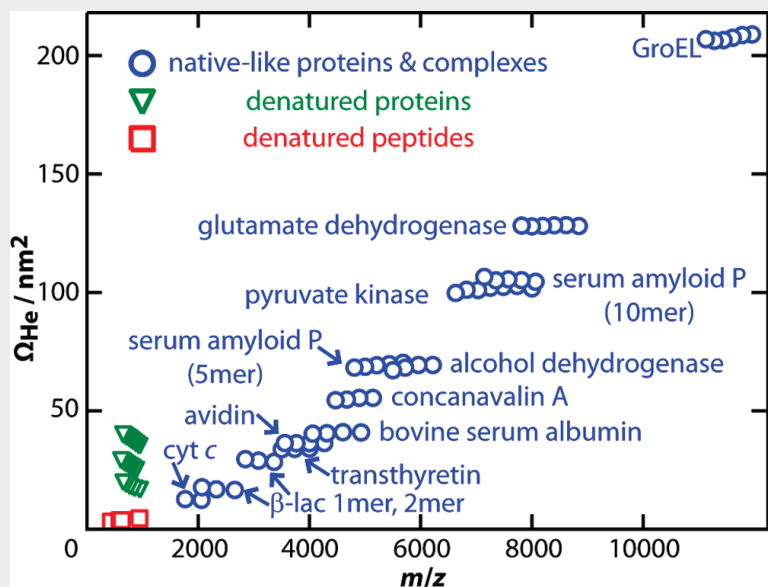
	m / Da	Z	$\Omega_{He} / \text{nm}^2$	$\Omega_{N_2} / \text{nm}^2$
GRGDS	490	1	1.32	2.06
		2	1.39	2.56
SDGRG	490	1	1.30	2.04
		2	1.42	2.59
Angiotensin fragment 1-7	898	2	2.26	3.34
RASG-1	1 000	2	2.25	3.31
Angiotensin II	1 046	2	2.45	3.35
Bradykinin	1 060	2	2.37	3.44
Angiotensin I	1 296	3	3.28	4.74
Renin substate	1 758	3	3.80	5.22
Enolase T35	1 872	3	3.80	5.19
Enolase T37	2 827	3	4.65	-

CCS values for proteins from TWIMMS (Waters)

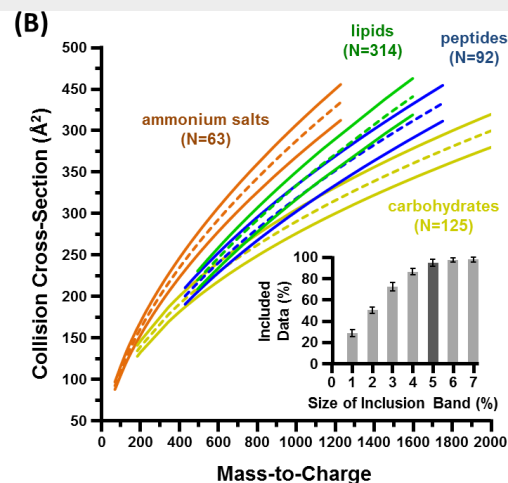
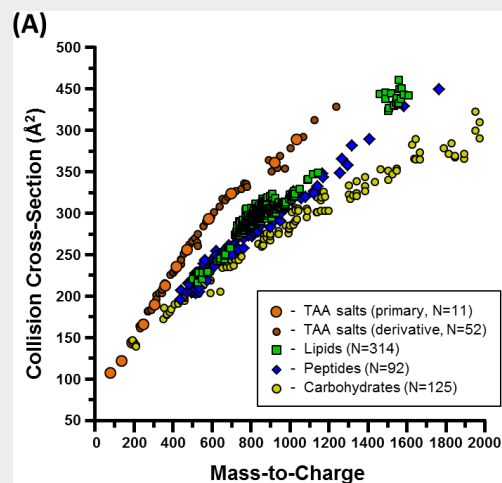
name		exact mass [Da]	CCS (this work ^a) [\AA^2]	CCS (literature ^b) [\AA^2]	abs. percent difference ^c [%]
tetramethylammonium	TAA1	74.14		107.40	
tetraethylammonium	TAA2	130.25		122.20	
tetrapropylammonium	TAA3	186.36	144.1 ± 0.7 (23)	143.80	0.22
tetrabutylammonium	TAA4	242.46	166.6 ± 0.9 (16)	166.00	0.36
tetrapentylammonium	TAA5	298.57	190.1 ± 1.0 (28)	190.10	0.02
tetrahexylammonium	TAA6	354.68	213.5 ± 1.0 (31)	214.00	0.23
tetraheptylammonium	TAA7	410.78	236.4 ± 0.4 (31)	236.80	0.17
tetraoctylammonium	TAA8	466.54	256.6 ± 0.7 (31)	258.30	0.64
tetradecylammonium	TAA10	579.11	293.5 ± 0.7 (24)		
tetradodecylammonium	TAA12	691.32	319.0 ± 0.9 (24)		
tetrahexadecylammonium	TAA16	915.04	361.5 ± 0.9 (24)		
tetraoctadecylammonium	TAA18	1027.16	379.0 ± 1.7 (21)		

CCS values for TAA salts from IM-Q-TOFMS (Agilent)

m/z vs. CCS Profiles for Different Classes



CCS values for proteins from TWIMMS (Waters)



CCS values for TAA salts from IM-Q-TOFMS (Agilent)

The Mobility Advantages

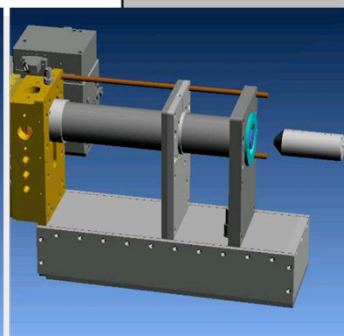
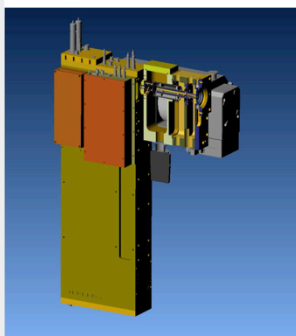
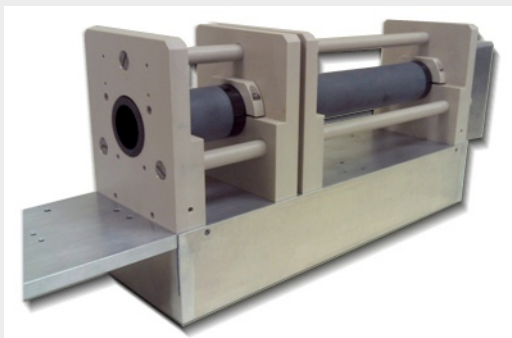
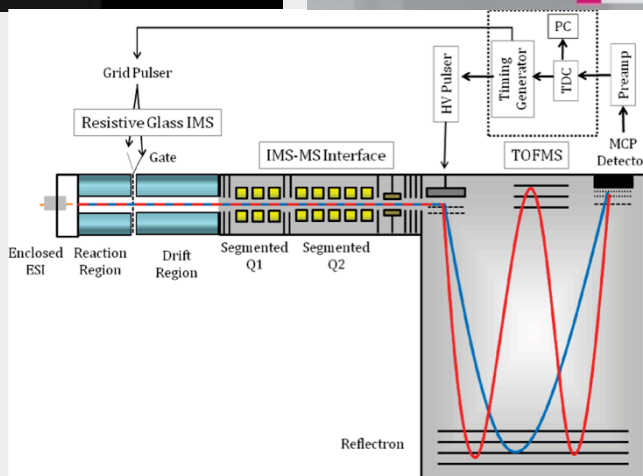


1. **Wide Application (metabolomics, glycomics, proteomics, etc.)**
2. **High Throughput Analysis (rapid preseparations)**
3. **Isomer Separation**
4. **Conformer Separation**
5. **Size Selective Fragmentation**
6. **Isotope Ratios in Complex Mixtures**
7. **Mobility-Mass Correlation for Chemical Classes**
8. **Surface analysis (image analysis)**
9. **Charge State Separation**
10. **Accurate Mobility Measurements**

11. **BUT THE MOST IMPORTANT MOBILITY ADVATAGE**



IMMS Instruments Are Now Available



QUESTIONS?



2014

ASMS, Baltimore, MD